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SYNTHESIS OF SOME 2'- AND 3'-FLUOROALKYL SUBSTITUTED NUCLEOSIDES AND OLIGONUCLEOTIDES

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As part of a programme on the development of antisense oligonucleotides, as selective inhibitors of oncogene expression, it was intended to investigate the properties of oligonucleotides modified at the 2'- or 3'-position with difluoromethylene, difluoromethyl and trifluoromethyl groups. It was expected that such oligonucleotides might possess increased stability against nucleases as well as improved hybridisation properties and transport characteristics (1,2,3).

The synthesis of target oligonucleotides entailed prior preparation of the appropriate nucleoside precursors. It was found that reaction of suitably protected 2'- and 3'-ketonucleosides with crystalline bromodifluoromethyl[tris (dimethylamino)]-phosphonium bromide in the presence of zinc gave 2'- and 3'-difluoromethylene nucleosides in high yields (4–6). Subsequently, these compounds were used as starting materials for further transformations.

Thus, hydrogenation of 2'-deoxy-2'-difluoromethylene-5'-O-dimethoxy-trityluridine (1) (4) and 3'-deoxy-3'-difluoromethylene-5'-O-dimethoxytrityluridine (4) (4), gave the corresponding 2'- and 3'-difluoromethyluridine derivatives 2a/2b (threo/erythro 6:1) and 5a/5b (threo/erythro 8:1), respectively. Detritylation of compounds 2a/2b and 5a/5b provided two pairs of diastereoisomers, 3a/3b (7) and 6a/6b (7), that could be separated by HPLC.

Interestingly, reaction of 2'-deoxy-2'-difluoromethylene-5'-O-dimethoxytrityl-3'-O-trimethylsilylethoxymethyluridine (11) (6) and 3'-deoxy-3' difluoromethylene-5'-O-dimethoxytrityl-2'-O-trimethylsilylethoxymethyluridine (17) (6) with tetrabutylammonium fluoride, resulted in fluorination at the unsaturated difluoromethylene carbon with loss of the trimethylsilylethoxymethyl

922

group and formation of 2',3'-didehydro-2',3'-dideoxy-5'-O-dimethoxytrityl-2'trifluoromethyluridine (13) and 2',3'-didehydro-2',3'-dideoxy-5'-O-dimethoxytrityl-3'-trifluoromethyluridine **19**, respectively.

No products of expected desilylation such as 12 and 18 were detected. Detritylation of 13 and 19 afforded the expected 2',3'-didehydro-2',3'-dideoxy-2' (3')trifluoromethyluridines 14 and 20. Hydrogenation of compounds 13 and 19 followed by detritylation provided 2',3'-dideoxy-2'-trifluoromethyluri dine (16a) and

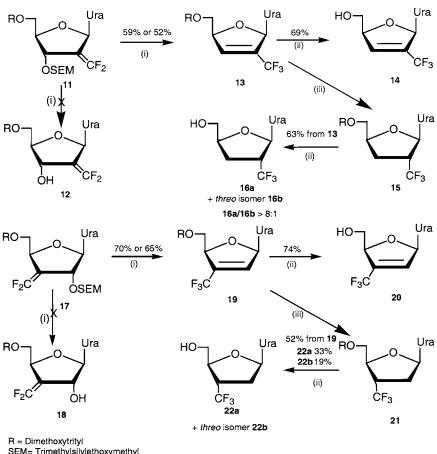
DMTrO Ura DMTrO Ura
$$\frac{73\%}{(ii)}$$
 DMTrO Ura $\frac{60\%}{(ii)}$ HO Ura $\frac{76\%}{3a}$ + erythro isomer 3b $\frac{60\%}{3a}$ + erythro isomer 3b $\frac{60\%}{3a}$ + erythro isomer 5b $\frac{60\%}{3a}$ + erythro isomer 6b $\frac{60\%}{3a}$ + erythro isomer 6

- Ura = Uracil-1-yl iPr = isopropyl
- (i) 10% palladium on activated carbon, EtOH; (ii) 80% aqueous acetic acid;
- (iii) diisopropylammoniumtetrazolide, NCCH2CH2OP(NiPr2)2, CH2Cl2.

Scheme 1. Synthesis of 3'- and 2'-O-Phosphoramidites of 2'- and 3'-Difluoromethyleneuridine and 2'- and 3'-Difluoromethyluridine.



2'- AND 3'-FLUOROALKYL SUBSTITUTED NUCLEOSIDES



SEM= Trimethylsilylethoxymethyl Ura = Uracil-1-vl

(i) Bu₄NF, THF, absence or presence of molecular sieves 3Å; (ii) 80% aqueous acetic acid; (iii) 10% palladium on activated carbon, EtOH.

Scheme 2. Synthesis of 2'- and 3'-trifluoromethyluridine derivatives.

2',3'-dideoxy-3'-trifluoromethyluridine (22a), along with the corresponding threo isomers 16b and 22b, respectively (6). (Scheme 2).

Finally, phosphitylation of compounds 1, 2a, 4 and 5a furnished the corresponding 2'- and 3'-O-phosphoramidites (7) (4), (8) (7), (9) (4) and (10) (7). (Scheme 1) Attempted incorporation of 3'-deoxy-3'-difluoromethylene-5'-O-dimethoxytrityluridine-2'-O-phosphoramidite (9) into oligonucleotide sequences was only possible after detailed studies in solution to customise the standard solid phase protocol. Replacement of iodine with t-butylhydroperoxide and the succinyl linker with oxalyl linker enabled the synthesis of short alternating oligonucleotides

The incorporation of 2'-deoxy-2'-difluoromethyl-5'-O-dimethoxytrityluridine-2'-O-phosphoramidite (8), and its 3'-difluoromethyl counterpart 10, required fewer changes in the standard protocol and resulted in (2'-5') and (3'-5') linked oligonucleotides modified with 3'- or 2'-difluoromethyl groups.

REPRINTS

ACKNOWLEDGMENTS

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- 7. All the NMR spectra were recorded in DMSO-d6 and UV spectra in 95% EtOH. 3a NMR $\delta_{\rm H}$ 3.02 (m, 1H, H-2'), 3.63 (m, 3H, H-4', H-5', H-5"), 4. 33 (t, 1H, H-3' J = 7.69 Hz), 4.95 (bs, 1H, 5'-OH), 5.61 (d, 1H, H-6, J = 8.12 Hz), 5.83 (d, 1H, 3'-OH, J = 7.25 Hz, 6.02 (t of d, 1H, CF₂H, $J_{HF} = 45.6 \text{ Hz}$, $J_{HH} = 4.47 \text{ Hz}$), 6.21 (d, 1H, H-1', J = 7.75 Hz) 7.84 (d, 1H, H-6, J = 8.12 Hz), 11.41 (bs, 1H, NH); δ_C 51.60 (t, $J_{C-F} =$ 19 Hz, C-2'), 57.96 (C-5'), 66.31 (C-3'), 81.04 (C-1'), 83.84 (C-4'), 100.44 (C-5), 114.54 (t, $J_{C-F} = 240$ Hz, CF_2 H), 140.18 (C-6), 149.31 (C-2), 162.13 (C-4); $\delta_F - 117.17$ $(1F, d (J_{FF} = 292 \text{ Hz}) \text{ of } d (J_{Hgem-F} = 54.6 \text{ Hz}), -121.46 (1F, d (J_{FF} = 292 \text{ Hz}) \text{ of } d$ $(J_{Hgem-F} = 53.53 \text{ Hz})$; UV λ_{max} 259 nm ε_{max} 6468, λ_{min} 229 nm ε_{min} 1564; Observed FAB MS 279.0770, $[C_{10}H_{12}F_2N_2O_5 + H]^+$ requires 279.0793. 3b NMR δ_H 2.89 (m, 1H, H-2'), 3.59 (m, 2H, H-5', H-5''), 3.85 (m, 1H, H-4'), 4.36 (d, 1H, -3', J = 1.50)4.48 Hz), 5.12 (bs, 1H, 5'-OH), 5.69 (d, 1H, H-5, J = 8.13 Hz), 5.79 (bs, 1H, 3'-OH), 6.33 (d, 1H, H-1', J = 8.53 Hz), 6.19 (t of d, 1H, CF_2H , $J_{HF} = 55.4 \text{ Hz}$, $J_{HH} =$ 6.80 Hz), 7.85 (d, 1H, H-6, J = 8.13 Hz), 11.32 (bs, 1H, NH); $\delta_F - 114.58$ (1F, d ($J_{FF} =$ 295 Hz) of q ($J_{Hgem-F} = 54.1$ Hz, $J_{H2'-F} = 10.0$ Hz), -123.51 (1F d ($J_{FF} = 295$ Hz) of q ($J_{Hgem-F} = 55.9 \text{ Hz}$, $J_{H2'-F} = 14.3 \text{ Hz}$); UV λ_{max} 260 nm ε_{max} 7426, λ_{min} 230 nm ε_{min} 1098; Observed FAB MS 279.0702, $[C_{10}H_{12}F_2N_2O_5^+H]^+$ requires 279.0793. **6a** NMR $\delta_{\rm H}2.87$ (m, 1H, H-3'), 3. 57 (m, 2H, H-5', H-5''), 4.30 (m, 2H, H-2', H-4'), 5.32 (bs, 1H, 5'-OH), 5.71(m, 3H, H-1', 2'-OH, H-5), 6.31 (t of d, 1H, CF_2H , $J_{HF} =$ 49.08 Hz, $J_{HH} = 6.88$ Hz), 7.84 (d, 1H, H-6, J = 8.16 Hz), 11.34 (bs, 1H, NHi);); δ_C C-2'), 49.38 (t, $J_{C-F} = 19.5 \text{ Hz}$, C-3'), 60.58 (C-5'), 76.90 (C-4'), 87.96 (C-1'), 102.40 (C-5), 116.96 (t, $J_{C-F} = 251$ Hz, CF_2H), 140.69 (C-6), 150.92 (C-2), 163.03 (C-4); δF -112.08 1F d ($J_{FF} = 294$ Hz) of q ($J_{Hgem-F} = 55.55$ Hz, $J_{H-3'-F} = 11.57$ Hz), -116.751F d ($J_{FF} = 294 \text{ Hz}$) of q ($J_{Hgem_F} = 56.3 \text{ Hz}$, $J_{H-3-F} = 15.08 \text{ Hz}$); UV λ_{max} 260 nm ε_{min} 8778 λ_{min} 229 nm ε_{min} 3436; Observed ES MS 279.0802, $[C_{10}H_{12}F_2N_2O_5 + H]^+$ requires 279.0793. **6b** NMR $\delta_{\rm H}$ 2.73 (m, 1H, H-3'), 3.52 (m, 1H, H-5'), 3.75 (m, 1H, H-5''), 4. 34 (m, 3H, H-2', H-4', 5'-OH), 5.61 (d, 1H, J = 8.24 Hz, H-5), 5.68 (bs, 2H, 3'-OH, H-1'), 6.20 (t, $J_{HF} = 56.1 \text{ Hz of d } J_{HH} = 5.28 \text{ Hz}$, 1H, CF_2H), 7.98 (d, 1H, H-6, J = 8.24 Hz), 11.30 (bs, 1H, NH); $\delta_F - 116.03 \text{ 1F d}$ ($J_{FF} = 290 \text{ Hz}$) of q ($J_{Hgem-F} =$



2'- AND 3'-FLUOROALKYL SUBSTITUTED NUCLEOSIDES

55.6 Hz, $J_{H3^-F}=10.06$ Hz) , -122.70 1F d ($J_{FF}=290$ Hz) of q ($J_{Hgem-F}=57.1$ Hz, $J_{H3'-F}=18.7$ Hz); UV λ_{max} 262 nm ε_{max} 8885 ε_{min} 230 nm ε_{min} 2020; Observed FAB MS 301.0600, [$C_{10}H_{12}F_2N_2O_5+Na]^+$ requires 301.0612. **3a** - 2D NOESY showed a cross peak between H-2' and H-4' and at the same time the lack of a cross peak between H-2' and H-5'. There was also a cross peak between H-6 and CF₂H. This is only possible if the H-2' proton is on the α face. **3b** - 2D NOESY showed a cross peak between H-2' and H-5 and at the same time the lack of a cross peak between H-2' and H-4'. This is only possible if the H-2' proton is on the β face. **6a** - 2D NOESY showed a cross peak between between H-5' and CF₂H. and at the same time the lack of a cross peak between H-3' and H-5'. There was also a cross peak between H-6 and CF₂H. This is only possible if the H-3' proton is on the α face. **6b** - 2D NOESY showed a cross peak between H-4' and CF₂H and at the same time the lack of a cross peak between H-5' and CF₂H. There was also a cross peak between H-3' and H-6. This is only possible if the H-3' proton is on the β face.

3. NMR $\delta_{\rm H}$ 1.05–1.14H (m, 14H, iPr), 2.59 (m, 1H, H-2'), 2.72 (t, 2H, J = 5.50 Hz, OCH₂CH₂CN), 3.45 (m, 2H, iPr), 3.51 (m, 2H, H-5', H-5''), 3.73 (s, 6H, OCH₃), 4.04 (m, 3H, OCH₂CH₂CN, H-4'), 4.67 (m, 1H, H-3'), 5.31 (d, 1H, H-5, J = 9.13 Hz), 6.10 t ($J_{\rm HF}$ = 56 Hz) of m, 1H, CF₂H), 6.30(m, 1H, H-1'), 6.87–7.42 (m, 13H, trityl), 7.65 (2 d, unresolved, H-6), 11.39 (bs, 1H, NH); $\delta_{\rm F}$ 117.9 (d of m, 1F, CF₂H), –122.2 (d of m, 1F, CF₂H) $\delta_{\rm P}$ 150.18 (s), 150.64 (s); Observed FAB MS 779.3050, [C₄₀H₄₇F₂N₄O₈P–H] requires 779.3021. **10** NMR $\delta_{\rm H}$, (1.21, m, 14H, iPr), 2.88 (t, 2H, J = 6.82 Hz, OCH₂CH₂CN), 2.99 (m, 1H, H-3'), 3.48(m, 4H, H-5', H-5'', iPr), 3.71 (s, 6H, OCH₃), 4.04 (m, 3H, OCH₂CH₂CN, H-4'), 4.51 (m, 1H, H-2'), 5.59 (d, 1H, H-5, J = 8.11 Hz), 5.89 (d, 1H, H-1', J = 5.62 Hz), 6.18 t ($J_{\rm HF}$ = 48.5 Hz) of m, 1H, CF₂H), 6.82–7.45 (m, 13H, trityl), 7.55 (d, 1H, H-6, J = 8.11 Hz), 11.34 (s, 1H, NH); $\delta_{\rm F}$ –112.58–(-114.86) (m, 1F, CF₂H) –117.11–(-118.99) (m, 1F, CF₂H); $\delta_{\rm P}$ 152.502 (s), 152.409 (s); Observed FAB MS 779.3063, [C₄₀H₄₇F₂N₄O₈P-H] requires 779.3021.

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